Enantioselective Preparation of β^2 -Amino Acids with Aspartate, Glutamate, Asparagine, and Glutamine Side Chains¹)

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(S)- β^2 -Homoamino acids with the side chains of Asp, Glu, Asn, and Gln have been prepared and suitably protected (N-Fmoc, CO₂/Bu, CONHTrt) for solid-phase peptide syntheses. The key steps of the syntheses are: N-acylation of 5,5-diphenyl-4-isopropyl-1,3-oxazolidin-2-one (DIOZ) with succinic and glutaric anhydrides ($Scheme\ 2$), alkylation of the corresponding Li-enolates with benzyl iodoacetate and Curtius degradation ($Scheme\ 4$), and removal of the chiral auxiliary ($Scheme\ 5$). In addition, numerous functional-group manipulations ($CO_2H \rightleftharpoons CO_2$ Bu, $CO_2Bn \rightleftharpoons CO_2H$, $CbzNH \rightarrow FmocNH$, $CO_2H \rightarrow CO_2NH_2 \rightarrow CONHTrt$; $Scheme\ 2$, 4, 5, and 6) were necessary, in order to arrive at the four target structures. The configurational assignments were confirmed by X-ray crystal-structure determinations ($Scheme\ 2$ and $Fig.\ 3$). The enantiomeric purities of a β^2h Asn and of a β^2h Gln derivative were determined by HPLC on a Chiralcel column to be 99.7:0.3 and SCHEM and SCHEM it took up to twelve steps to prepare a suitably protected trifunctional product with a single stereogenic center (overall yield of 10% from DIOZ and succinic anhydride)!

1. Introduction. – Although β^3 -amino acids can be conveniently synthesized from the corresponding α -analogues by simply introducing a CH₂ group between the C=O and the α -C-atom (i.e., Arndt-Eistert homologation)⁵), no such general method is known for the preparation of β^2 -amino acids. As outlined in Fig. 1, a wide variety of routes have been explored for the synthesis of β^2 -amino acids depending upon the bond formed in the key step.

Formation of bond ① by diastereoselective *Mannich*-type reaction [1-11], *i.e.*, aminomethylation of chiral acyl derivatives with synthetic equivalents of the $[H_2NCH_2]^+$ cation, has been applied for the preparation of a large range of β^2 -amino acids, most of them having alkyl or aryl side chains. The *Mitsunobu* reaction, instead, was used for the conversion of 2-(hydroxymethyl)carboxylic acid esters to the corresponding amino or azido derivatives, thus forming bond ② [1][12][13]. Another route is the *Curtius* rearrangement of enantiomerically pure monosubstituted succinic acid derivatives [1][14][15], which were themselves synthesized by alkylation of chiral

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⁵⁾ With the exceptions of Fmoc-β³hHis(Trt)-OH and Fmoc-β³hCys(Trt)-OH, all 'proteinogenic' β³-amino acids, which can be prepared by Arndt-Eistert homologation of the corresponding α-amino acids, are commercially available.

Fig. 1. Analysis of the key C-C-, C-N-, or C-H-bond-forming reactions for β^2 -amino acid synthesis

N-acyloxazolidinones with bromo- or iodoacetates (bonds ① and ②) [9][16–18] or by alkylation of succinic [19] or malic acid enolates [20] with electrophiles (bonds ③ and ②). Another approach is the coupling of an electrophile with cyclic [21–28] or acyclic [1][8][29–33] chiral enolates derived from 3-aminopropanoic acid (bond ③). Similarly, by the enantioselective nucleophilic addition of Et_2Zn to a 3-nitroacrylate derivative, the feasibility of the process with *umpolung* has been demonstrated [34]. The conjugate 1,4-addition (*Michael*-type reaction) of chiral N-nucleophiles to 2-substituted acrylates, followed by diastereoselective protonation, is yet another method for their preparation, in which bonds ② and ④ are formed [35–38]. Finally, chiral metal complexes have permitted the isolation of enantiomerically enriched β^2 -amino acids by construction of the ① + ④ [39], ④ + ⑤ [40], or ④ + ⑦ bonds [41], albeit often with low enantioselectivities and, in any case, applied only for derivatives with alkyl or aryl side chains.

The incorporation of β^2 -amino acids into β -peptides has uncovered intriguing effects, leading to turn structures [42][43] or a novel type of helix, which consists of alternating ten- and twelve-membered H-bonded rings [7][44]. For our ongoing work on the synthesis of more complex β^2/β^3 - and all- β^2 -peptides, we needed ample access to all 19, appropriately protected⁶), β^2 -amino acids with proteinogenic side chains. At the time this project was initiated, the above mentioned methods were only applied to the construction of the 'easy' β^2 -amino acids, but not much was known about the derivatives with functionalized side chains. For this reason, the application of a method developed in our group, relying on the use of 5,5-diphenyl-4-isopropyl-1,3-oxazolidin-2-one (DIOZ) as a chiral auxiliary, was envisaged for the preparation of the β^2 -amino acids shown in *Fig.* 2. Indeed, our modified *Evans* oxazolidinone has been shown to be a versatile auxiliary for diastereoselective alkylations, aldol additions, 1,4-additions, cycloadditions, and numerous other reactions [8][45–53]. Derivatives of this auxiliary have turned out to possess several desirable properties, such as high crystallinity, ease of removal and recovery, and good selectivity.

Results and Discussion. – Since the aspartic and the glutamic acid side chains are interchangeable with the corresponding amides (asparagine and glutamine, resp.), and since they differ from each other only by a CH_2 unit, it was decided to prepare all four β^2 -amino acids by the same approach. As mentioned above, a prerequisite of the final

⁶⁾ For solid-phase synthesis by the Fmoc strategy, all functionalized side chains have to be protected orthogonally to the Fmoc group itself. They should be stable under basic conditions but labile in acidic media, thus trityl (Trt) and 'Bu groups are the protecting groups of choice.

Fig. 2. Target β^2 -amino acid derivatives

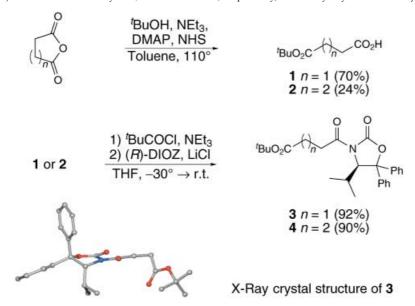
compounds for their use in solid-phase β -peptide synthesis is the Fmoc protection of the β -amino group, and the 'Bu or Trt protecting group on the side chains. Thus, we thought that 3-acyl-DIOZ derivatives of succinic and glutaric acids could serve as precursors for the corresponding β^2 -homoaspartic and β^2 -homoglutamic acids, respectively. The amino functionality would be introduced either by a diastereoselective *Mannich*-type reaction (*cf.* bond ① in *Fig.* 1) or by a carbalkoxymethylation, followed by *Curtius* degradation (connections ① + ② in *Fig.* 1), to give the desired amino acid skeleton, which could be further elaborated by protecting-group exchange (*Scheme* 1).

Scheme 1. Retrosynthetic Analysis for the Preparation of the β^2 -Amino Acids with Asx and Glx Side Chains

XOC
$$n$$
 OH $n = 1, 2$ $n = 1, 2$

It should be noted that a similar approach has been previously described by Wyatt and co-workers [15] for the synthesis of fully deprotected β^2 -homoaspartate; the authors applied a diastereoselective alkylation of a 'classical' Evans acyloxazolidinone derived from pent-4-enoic acid with methyl bromoacetate as electrophile, followed by oxidative cleavage of the double-bond, Curtius rearrangement, and removal of the auxiliary, to give the Z- β^2 hAsp-OH in 97.5% enantioselectivity, but without a protecting group on the side chain. Recently, Xue et al. [32] performed the synthesis of an enantiomerically pure (R)- β^2 -homoaspartate derivative by a stereoselective alkylation of a Bn- and Z-protected chiral β -homoglycine-oxazolidinone with tert-butyl bromoacetate. In our approach, we started from the commercially available succinic and glutaric anhydrides, which were ring-opened by refluxing in toluene with 'BuOH, and catalytic amounts of 4-(dimethylamino)pyridine (DMAP), Et₃N, and N-hydroxysuccinimide (NHS) [54] to give the monoesters 1 and 2 in 70 and 24% 7) yield, respectively (Scheme 2). N-Acylation of the chiral oxazolidinone (R)-DIOZ with the LiCl-activated [55] pivaloyl mixed anhydrides of 1 and 2 gave the desired acyloxazolidinones 3 and 4, respectively, in excellent yields.

Scheme 2. Nucleophilic Ring Opening of the Succinic and Glutaric Anhydrides ($\rightarrow 1$ and 2), N-Acylation of (R)-DIOZ to Form the Acyloxazolidinones 3 and 4, Respectively, and X-Ray-Crystal Structure of 3



Slow evaporation of an Et_2O /pentane mixture from a solution of ester 3 led to the formation of crystals suitable for structure determination by single-crystal X-ray diffraction; as evident from the presentation in *Scheme 2*, the methine H-atom of the 1 Pr group points towards the *cis*-Ph group causing a buttressing effect [49] in the face selectivity of reactions of enolates of 3 with electrophiles.

⁷⁾ The reaction was rather messy and the formation of polymerization products were observed.

Due to the better crystallinity of compound 3 and, therefore, easier purification of its derivatives, compared to the homologue 4, all experiments towards the construction of the amino acid backbone, according to the approaches outlined in Scheme 1, were performed with the former derivative. We first explored amidomethylation of the Tienolate from 3 with the corresponding electrophile (N-benzyloxycarbonyl-N-(methoxymethyl), ZNHCH2OMe) without success; the Lewis acidity of Ti4+ caused complete deprotection of the tert-butyl ester and formation of an inseparable mixture of products. We then switched to an allylation/double-bond oxidation/Curtiusdegradation strategy. An analogous stereoselective allylation reaction has been successfully applied by Sibi and Deshpande [19] on a similar substrate. Treatment of 3 with 1 equiv. of sodium hexamethyldisilazide (NHMDS) in THF at -78° for 20 min, followed by quenching with allyl bromide, furnished, in our case, the mono alkylated derivative 5 in disappointing yields and selectivities (Entry 1, Scheme 3). Similar results were obtained when more equivalents of allyl bromide were added to the Na-enolate (Entry 2); however, better conversion was achieved when the more reactive iodo derivative was used as electrophile (Entry 3). Acceptable selectivities were realized only by running the reaction at -50° (Entry 4), while complete degradation of the enolate resulted at 0° (Entry 5). The formation of the Li-enolate with LHMDS at -78° ,

Scheme 3. Optimization of the Conditions for the Diastereoselective Alkylation of 3 with Allyl Halides

$$t_{BuO_2C}$$
 t_{BuO_2C}
 t_{BuO_2C}

Entry	Base, equiv.	C_3H_5X , equiv.	Time [h]	Temp. [°]	Conversion [%] ^a) d.r. ^a)
1	NHMDS, 1.1	X = Br, 1.5	14	-78 → -48	3 41	75:25
2	NHMDS, 1.1	X = Br, 5	22	-78	43	84:16
3	NHMDS, 1.1	X = 1, 5	6	-78	85	80:20
4	NHMDS, 1.1	X = 1, 5	6	$-78 \rightarrow -50$	88	94:6
5	NHMDS, 1.1	X = 1, 5	6	$-78 \rightarrow 0$	_b)	_c)
6	KHMDS, 1.1	X = 1, 5	15	$-78 \rightarrow -50$) _b)	-c)
7	LHMDS, 1.1	X = I, 5	23	-78 → -50	85	96:4
8	LHMDS, 1.5	X = 1, 5	20	$-78 \rightarrow -50$	81	94:6
9	LHMDS, 2	X = 1, 5	20	$-78 \rightarrow -50$	69	-c)

a) Determined by ¹H-NMR spectroscopy of the crude product. ^b) Enolate degradation and formation of the allylated auxiliary as major product. ^c) Not determined.

followed by quenching with allyl iodide at -50° , gave the best result, with good conversion and 96% diastereoselectivity (*Entry 7*). Unfortunately, purification by flash chromatography was not sufficient to separate the diastereoisomers, and compound **5** was isolated in 66% yield and 96% diastereoselectivity. At this point, we thought that better separation would possibly be achieved in a later stage of the synthesis, for example, after double-bond oxidation. Ozonolysis [56] of the double bond in **5** and oxidation with periodate (catalytic KMnO₄ [57]) did, however, not lead to the desired carboxylic acids.

Disappointed by these results, we changed our strategy and envisaged a more direct approach of introducing the amino functionality by alkylation with benzyl iodoacetate [58] as electrophile. We were pleased to see that, upon treatment of the Li-enolates of the acyloxazolidinones 3 and 4 with this electrophile at -78° , the alkylated products 6 and 7, respectively, could be isolated in *ca.* 55% yield as single diastereoisomers (*Scheme 4*).

Scheme 4. Diastereoselective Alkylation of Succinic and Glutaric Acid Derivatives 3 and 4 with Benzyl Iodoacetate. Note that the α-carbonyl C-atom of 6 is a stereogenic center only due to the fact that the two ester groups CO₂R carry different OR groups ('Bu and Bn)! Hydrogenolysis of the benzyl esters 6 and 7, and Curtius degradation leads to the Z-protected derivatives 10 and 11.

Following hydrogenolysis of the benzyl ester and *Curtius* degradation (with DPPA (diphenylphosphoryl azide), Et_3N , and BnOH [59]), the Z-protected derivatives **10** and **11** were isolated in good yields (*Scheme 4*). Compound **10** formed single crystals suitable for structure determination by X-ray crystallography (*Fig. 3,a*): as expected, the newly created stereogenic center has (*S*)-configuration. In addition, X-ray crystal-structure determination of the fifth crop of crystals (representing less than 4% of the total yield) revealed that the unit cell contains **10** and *epi-10* in a 1:1 ratio (*Fig. 3,b*).

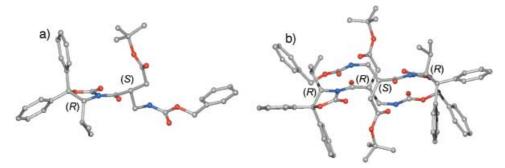


Fig. 3. X-Ray crystal structures of 10 (a) and of a 1:1 co-crystal of 10 and epi-10 (b)

The next step towards the synthesis of the β^2 -aspartate and β^2 -glutamate derivatives involved the cleavage of (R)-DIOZ with LiOH/H₂O₂⁸), which gave acids **12** and **13**, as well as the recovered auxiliary, in good yields $(Scheme\ 5)^9$). Following Z/Fmoc protecting-group exchange, Fmoc- β^2 hAsp(O'Bu)-OH (**14**) and Fmoc- β^2 hGlu(O'Bu)-OH (**15**) were isolated in 94 and 90% yields, respectively. In this way, the Fmoc derivatives **14** and **15** were synthesized in an overall yield of 19 and 7% ¹⁰) over eight steps starting from the corresponding anhydrides and (R)-DIOZ.

Scheme 5. Cleavage⁸) of the Auxiliary with LiOH/H₂O₂ and Subsequent Protecting-Group Exchange to Give the Fmoc-Protected Acids **14** and **15**

10 or THF/H₂O, 0°, 1.5 h
$$^{t}BuO_{2}C$$
 $^{t}DuO_{2}C$ ^{t}DuO

For the preparation of the β^2 hAsn and β^2 hGln derivatives, the acids **12** and **13** were converted to the benzyl esters **16** and **17**, respectively, by treatment with BnBr and NaHCO₃ in DMF in the presence of catalytic amounts of tetrabutylammonium iodide (TBAI; *Scheme* 6). It is important to note that nucleophilic cleavage of the acyloxazolidinone **10** by BnOLi, for the direct formation of ester **16**, led to partial epimerization of the C(2)-centre (see above). Subsequent 'Bu deprotection and nucleophilic addition of ammonia afforded amides **18** and **19**, which were Trt-protected

⁸⁾ We have claimed that a major advantage of our modified Evans auxiliary is that it can be cleaved simply with THF/aq. NaOH and does not require LiOH/H₂O₂. This is not true for the application described here, because the substrate contains ester groups, and the hydroperoxide conditions lead to much faster cleavage.

⁹⁾ Particular care should be taken during the workup, the tert-butyl ester being very sensitive to acidic conditions. For details, see Exper. Part.

¹⁰⁾ The low overall yield for this derivative is caused 'essentially' by the disappointing yield obtained for the opening of the glutaric anhydride (not optimized).

to give the fully protected amino acids **20** and **21**, respectively, in good yields. Hydrogenolysis and Fmoc protection of the amino group led to Fmoc- and Trt-protected β^2 -homoasparagine (**22**) and β^2 -homoglutamine (**23**; *Scheme 6*), respectively, which have, thus, been synthesized over twelve steps with an overall yield of 10 and 4%, respectively.

Scheme 6. Conversion of the Aspartate and Glutamate Derivatives 12 and 13 to Fmoc- and Trt-Protected β^2 Homoasparagine (22) and β^2 -Homoglutamine (23)

12 or BnBr, NaHCO₃ Bu₄NI DMF, r.t., 20 h

13
$$\frac{1}{13}$$
 $\frac{1}{13}$ $\frac{1}$

To verify the enantiomeric purity of the new β^2 -amino acid derivatives, compounds **20** and **21** were injected into a chiral HPLC column (*Chiralcel OD*). For comparison, both derivatives were racemized by treatment with BnOLi at room temperature. As shown in *Fig. 4*, both derivatives show only one peak, thereby confirming the purity of all four amino acids¹¹). Additionally, as previously mentioned, direct cleavage of the auxiliary from acyloxazolidinone **10** with BnOLi (1 equiv. BuLi, 0° , 4 h) gave ester **16**. This compound was further derivatized to give ester (*R*)-**20**, the enantiomeric purity of which was determined to be 66% by HPLC. Since this alternative route differs from that outlined in *Scheme 3* only in the reaction involving the removal of the auxiliary, we can conclude that the cleavage of the auxiliary with BnOLi is not always free of epimerization/racemization⁸).

In conclusion, we have shown the remarkable potential of 3-acyl-DIOZ derivatives for the preparation of β^2 -amino acids carrying the side chains of Asp, Asn, Glu, and Gln. Furthermore, we have judiciously verified the enantiomeric purity of these compounds by HPLC on a chiral stationary phase in order to ensure their utility as building blocks for solid-phase peptide synthesis according to the Fmoc protocol. Our approach can easily be scaled up to produce multigram amounts of these amino acids in enantiomerically pure form.

⁽S)-form of the asparagine, but the (R)-form of the glutamine derivative, is eluted first.

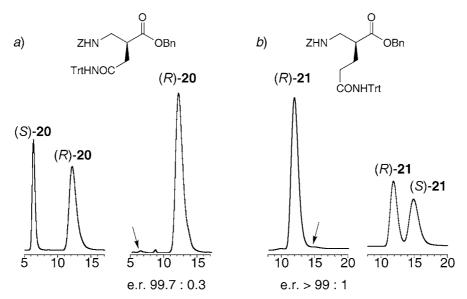


Fig. 4. HPLC Traces of a) the asparagine derivative (R)-20 and of rac-20, and b) the glutamine derivative (R)-21 and rac-21 (Chiralcel OD-H, λ = 220 nm, mobile phase: PrOH/hexane 20:80 for 20 and PrOH/hexane 15:85 for 21). The arrows indicate the signals of the minor enantiomers. The e.r. values were determined by integration of the corresponding peaks: (R)-20:99.7:0.3; (R)-21: >99:1. t_R in min.

Experimental Part

- 1. General. Abbreviations: CDI: 1,1'-Carbonyldi(1H-imidazole), DIOZ: 5,5-diphenyl-4-isopropyl-1,3-oxazolidin-2-one, DMAP: 4-(dimethylamino)pyridine, DPPA: diphenylphosphoryl azide, FC: flash chromatography, h.v.: high vacuum, 0.01–0.1 Torr, LHMDS: lithium hexamethyldisilazide, NHMDS: sodium hexamethyldisilazide, TBAI: tetrabutylammonium iodide, TES: Et₃SiH. Solvents for chromatography and workup procedures were distilled from Sikkon (anh. CaSO₄; Fluka), THF was distilled from Na, CH₂Cl₂ and Et₃N from CaH₂. LiCl was dried in h.v. at 100° for 1 h. All other reagents were used as received from Fluka. TLC: Merck silica-gel 60 F₂₅₄ plates; detection with UV or 'Mo-stain' soln. (25 g phosphormolybdic acid, 10 g Ce(SO₄)₂·H₂O, 60 ml conc. H₂SO₄, 940 ml H₂O). FC: Fluka silica gel 60 (40–63 μm); at ca. 0.2 bar. Anal. HPLC: Waters HPLC system (pump type 515, data module type 746, tunable absorbance detector type 484). Chiralcel OD-H column. M.p.: Büchi-510 apparatus; uncorrected. Optical rotations: Perkin-Elmer 241 polarimeter (10 cm, 1-ml cell). IR Spectra: Perkin-Elmer 1600 FT-IR spectrophotometer. NMR Spectra: Bruker AMX-400 (¹H 400 MHz, ¹³C 100 MHz); chemical shifts δ in ppm downfield from internal Me₄Si (=0 ppm); J values in Hz. Mass Spectra: IonSpec Ultima 4.7 T FT ion cyclotron resonance (ICR, HR-MALDI, in 2,5-dihydroxybenzoic acid matrix) spectrometer; in m/z (% of basis peak). Elemental analyses were performed by the Microanalytical Laboratory of the Laboratorium für Organische Chemie, ETH-Zürich.
- 2. Preparation of Diacid Mono-tert-butyl Esters: General Procedure 1 (GP 1). Similarly to the procedure reported in [54], the corresponding anhydride was suspended in toluene (1.7M) under Ar, and treated subsequently with N-hydroxysuccinimide (0.3 equiv.), DMAP (0.1 equiv.), dry 'BuOH (3 equiv.) and Et₃N (0.3 equiv.). The mixture was stirred for 30 min at r.t., and heated at reflux for 1 d. After cooling, AcOEt was added, the org. phase washed with 10% citric acid (3×) and sat aq. NaCl soln. (1×), dried (MgSO₄), and concentrated under reduced pressure. FC or recrystallization yielded the pure product.
- 3. Acylation of the Auxiliary by the Mixed Anhydride Method: General Procedure 2 (GP 2). Similarly to the procedure reported in [8], to a soln. of the corresponding carboxylic acid in THF (0.2M) under Ar, cooled at -30° , Et₃N (2.5 equiv.) and pivaloyl chloride (1 equiv.) were added dropwise. The resulting suspension was stirred at -30° for 2 h. At this point, LiCl (1.1 equiv.) and the auxiliary (0.95 equiv.) were added at once, stirring was continued for 1-2 d as the mixture was slowly allowed to warm to r.t. The mixture was diluted with Et₂O,

washed with sat. aq. NH_4Cl soln. (2×), 1M NaOH (2×), and sat. aq. NaCl soln., dried (MgSO₄), and concentrated under reduced pressure. FC of the crude product yielded the pure compound.

- 4. Alkylation of Acyloxazolidinones with Benzyl Iodoacetate: General Procedure 3 (GP 3). To a soln. of the corresponding substrate in THF (0.3M) at -78° , LHMDS (1.1 equiv.) was slowly added, and the mixture was stirred for 1 h at this temp. At this point, a soln. of benzyl iodoacetate (1.5 equiv.) in THF (1:1, v/v) was slowly added over 1 h, and the resulting mixture was stirred for 20 h at -50° . The mixture was then diluted with Et₂O and washed with sat. aq. NH₄Cl soln., 5% NaHSO₃, and sat. aq. NaCl soln. The org. phase was dried (Na₂SO₄), and concentrated under reduced pressure. FC of the crude product yielded the pure compound.
- 5. Selective Hydrogenolysis of the Bn Ester in the Presence of DIOZ: General Procedure 4 (GP 4). The corresponding substrate was dissolved in THF (0.02M) and ca. 10% (w/w) Pd/C (10%) was added. The apparatus was evacuated and flushed with H₂ (3 \times), and the soln. stirred under an atmosphere of H₂ (balloon) for 1.5 h (TLC control). The mixture was then filtered through Celite and washed thoroughly with CH₂Cl₂. The filtrate was collected and concentrated under reduced pressure. FC of the crude product yielded the pure compound.
- 6. Curtius *Rearrangement: General Procedure 5 (GP 5)*. The corresponding substrate was dissolved in toluene (0.15m) under Ar, and Et₃N (1.2 equiv.), followed by DPPA (1.2 equiv), was added. The resulting mixture was stirred for 30 min at r.t., and subsequently heated at reflux for 1.5 h. BnOH (2 equiv.) was then added, and the mixture was refluxed for further 16 h. After cooling, the mixture was diluted with Et₂O, washed with sat. aq. NaHCO₃ soln. and sat. aq. NaCl soln., dried (NaSO₄), and concentrated under reduced pressure. FC of the crude product yielded the pure compound.
- 7. Cleavage of the Chiral Auxiliary with LiOH/ H_2O_2 : General Procedure 6 (GP 6). To a soln. of the corresponding substrate in THF/ H_2O 4:1 (0.2M) at 0°, H_2O_2 (30% aq., 4.0 equiv.) and LiOH· H_2O (1.6 equiv.) were added, and the resulting mixture was stirred at 0° for 2 h. The mixture was diluted with H_2O , and the precipitated chiral auxiliary was removed by filtration. The THF was evaporated at 25° under reduced pressure, the soln. was cooled to 0°, and AcOEt was added to the mixture. Upon slow addition of 10% citric acid and with continuous stirring of the biphasic system at 0°, the pH of the aq. phase was adjusted to 4–5. The org. layer was separated, and the aq. phase was extracted with AcOEt (3×). The combined org. layers were washed with sat. aq. NaCl soln., dried (Na₂SO₄), and concentrated under reduced pressure. FC of the crude product yielded the pure compound.
- 8. Hydrogenolysis of Z groups and Bn Esters: General Procedure 7 (GP 7). The corresponding substrate was dissolved in MeOH (0.02M), and ca. 10% (w/w) Pd/C (10%) was added. The apparatus was evacuated and flushed with H_2 (3 \times), and the soln. was stirred under an atmosphere of H_2 (balloon) for 1–2 h (TLC control). Subsequent filtration through Celite and concentration under reduced pressure yielded the crude product, which was used in the next step without further purification.
- 9. Fmoc Protection: General Procedure 8 (GP 8). The N-deprotected amino acid was dissolved in 0.15m Na₂CO₃ (2–3 equiv.) and treated with a soln. of FmocOSu (1–1.2 equiv.) in acetone (0.1m). If necessary the pH was readjusted to 9–10 with additional aq. Na₂CO₃ soln., and the mixture was stirred at r.t. for 4 h. The acetone was carefully removed under reduced pressure at 30°, and the resulting mixture was diluted with H₂O. At this point, the pH of the soln. was adjusted to 9–10 with 0.6m aq. Na₂CO₃. The aq. mixture was then extracted with Et₂O (2×). The aq. phase was separated, cooled to 0°, and AcOEt was added to the mixture. With continuous stirring of the biphasic system at 0°, the pH of the aq. phase was adjusted to 4–5 by slow addition of 10% aq. citric acid. The org. layer was separated, and the aq. layer was extracted with AcOEt (2×). The combined org. layers were washed with sat. aq. NaCl soln., dried (Na₂SO₄), and concentrated under reduced pressure. FC of the crude product yielded the pure compound.
- 10. Preparation of Bn Esters: General Procedure 9 (GP 9). To a soln. of the corresponding acid in DMF (0.2M) were added NaHCO₃ (2.0 equiv.), TBAI (0.3 equiv.), and BnBr (5.0 equiv.). The resulting mixture was vigorously stirred at r.t. for 20 h. The mixture was diluted with Et₂O, and the org. phase was washed with H₂O and sat. aq. NaCl soln. The org. phase was dried (Na₂SO₄) and concentrated under reduced pressure. FC of the crude product yielded the pure compound.
- 11. 1 Bu Ester Deprotection: General Procedure 10 (GP 10). To a soln. of the corresponding substrate in CH₂Cl₂ (0.4M), TES (2.5 equiv.) and TFA (13.0 equiv.) were added dropwise, and the mixture was stirred at r.t. for 2 h. The solvent was removed under reduced pressure at 25° and residual TFA co-evaporated with toluene (3×). The product obtained was used in the next step without further purification.
- 12. Conversion of an Acid to an Amide: General Procedure 11 (GP 11). The corresponding acid was dissolved in THF (0.13m), and CDI (2 equiv.) was added. After stirring for 5 h under Ar, excess NH₃ gas was bubbled through the soln. The mixture was then diluted with AcOEt, and the org. phase was washed with H₂O,

1 M HCl, and sat. aq. NaCl soln. The org. phase was dried (Na_2SO_4) and concentrated under reduced pressure. Crystallization yielded the pure compound.

13. N-Trt Protection: General Procedure 12 (GP 12). To a suspension of the corresponding substrate in AcOH (0.27M), solid TrtOH (2.0 equiv.) was added, followed by a soln. of Ac₂O/AcOH/H₂SO₄ 100:200:3.6 (0.56 ml/mmol). The mixture was heated at 40° and stirred for 5.5 h. Et₂O was then added, and the org. phase was washed with 5% NaHCO₃ and sat. aq. NaCl soln. The org. phase was dried (Na₂SO₄) and concentrated under reduced pressure. FC of the crude product yielded the pure compound.

4-(tert-Butoxy)-4-oxobutanoic Acid (1). Transformation of succinic anhydride (30 g, 0.3 mol) according to GP1, followed by recrystallization (Et₂O/pentane), afforded 1 (36.36 g, 70%). White solid. The anal. data are in agreement with the values reported in [54][60].

5-(tert-Butoxy)-5-oxopentanoic Acid (2). Transformation of glutaric anhydride (34.2 g, 0.3 mol) according to $GP\ 1$, followed by FC (Et₂O/pentane 1:1), afforded 2 (13.6 g, 24%). Colorless oil. The anal. data are in agreement with the values reported in [61].

(R)-3-{3-[(tert-Butoxy)carbonyl]-1-oxopropyl]-4-(1-methylethyl)-5,5-diphenyloxazolidin-2-one (3). Reaction of 1 (6.0 g, 34.4 mmol) with (R)-DIOZ (9.21 g, 32.7 mmol) according to GP 2, followed by FC (Et₂O/pentane 1:5), afforded 3 (13.2 g, 92%). White solid. M.p. $149-151^{\circ}$. $R_{\rm f}$ (Et₂O/pentane 1:5) 0.30. [a]_D^{1.1} = +184.0 (c = 1.00, CHCl₃). IR (CHCl₃): 2978w, 1782s, 1718m, 1494w, 1450w, 1368m, 1320w, 1153m, 1051w, 1001w, 941w, 845w. ¹H-NMR (400 MHz, CDCl₃): 0.77 (d, J = 6.8, Me); 0.87 (d, J = 7.0, Me); 1.40 (s, 'Bu); 1.90 – 2.00 (m, Me₂CH); 2.43 – 2.59 (m, CH₂); 2.93 (ddd, J = 12.3, 6.8, 5.4, 1 H, CH₂); 3.26 (ddd, J = 13.9, 8.2, 6.7, 1 H, CH₂); 5.38 (d, J = 3.3, CHN); 7.26 – 7.40 (m, 8 arom. H); 7.45 – 7.48 (m, 2 arom. H). ¹³C-NMR (CDCl₃, 100 MHz): 16.32, 21.72, 28.00 (Me); 29.77 (CH₂); 30.01 (CH); 30.88 (CH₂); 64.49 (CH); 80.46, 89.46 (C); 125.68, 125.96, 128.37, 128.61, 128.92 (CH); 138.24, 142.32, 152.99, 171.32, 171.93 (C). MALDI-MS: 460.2 (44.34), 404.1 (100.00). Anal. calc. for C₂₆H₃₁NO₅ (437.53): C 71.37, H 7.14, N 3.20; found: C 71.40, H 7.26, N 3.15.

(R)-3-[4-[(tert-Butoxy)carbonyl]-1-oxobutyl]-4-(1-methylethyl)-5,5-diphenyloxazolidin-2-one (4). Reaction of **2** (6.5 g, 34.5 mmol) with (R)-DIOZ (9.23 g, 32.8 mmol) according to GP 2, followed by FC (Et₂O/pentane 1:5), afforded **4** (13.3 g, 90%). White solid. M.p. 92–93°. $R_{\rm f}$ (Et₂O/pentane 1:5) 0.30. [a] $_{\rm b}^{\rm L}$ = +173.0 (c = 1.00, CHCl₃). IR (CHCl₃): 3011w, 2972w, 1781s, 1718s, 1494w, 1450m, 1368m, 1320w, 1320m, 1154m, 1050w, 1001w, 844w, 638w. $^{\rm H}$ -NMR (400 MHz, CDCl₃): 0.76 (d, J = 6.8, Me); 0.87 (d, J = 70, Me); 1.42 (s, $^{\rm B}$ u); 1.78 – 2.01 (m, CH₂), Me₂CH); 2.20 (t, J = 7.4, CH₂); 2.77 (ddd, J = 17.0, 8.0, 6.6, 1 H, CH₂); 2.96 (ddd, J = 17.0, 8.2, 6.6, 1 H, CH₂); 5.37 (d, J = 3.4, CHN); 7.25 – 7.40 (m, 8 arom. H); 7.45 – 7.48 (m, 2 arom. H). 13 C-NMR (100 MHz, CDCl₃): 16.40 (Me); 19.91 (CH₂); 21.77, 28.07 (Me); 29.85 (CH); 34.31, 34.48 (CH₂); 64.47 (CH); 80.27, 89.40 (C); 125.58, 125.92, 127.94, 128.36, 128.60, 128.90 (CH); 138.14, 142.29, 152.98, 172.20, 172.42 (C). MALDI-MS: 474.2 (46.16), 418.2 (98.46). Anal. calc. for C₂₇H₃₃NO₅ (451.56): C 71.83, H 7.37, N 3.10; found: C 71.81, H 7.24, N 3.36

(R)-3-((R)-2-{[(tert-Butoxy)carbonyl]methyl}-1-oxopent-4-enyl)-4-(1-methylethyl)-5,5-diphenyloxazolidin-2-one (**5**). Compound **3** (150 mg, 0.34 mmol) was dissolved in THF (0.1M) and cooled to -78° . LHMDS (1M in THF, 0.38 ml, 1.1 equiv.) was added dropwise, and the mixture was stirred for 1.5 h at this temp. At this point, allyl iodide (0.16 ml, 5 equiv.) was added slowly over 30 min, and the resulting mixture was stirred at -50° for 23 h. The mixture was quenched with sat. aq. NH₄Cl soln. and diluted with Et₂O. The org. phase was washed with 1M HCl, 1M NaOH, and sat. aq. NaCl soln., dried (MgSO₄), and concentrated under reduced pressure. FC (Et₂O/pentane 1:6) of the crude product yielded **5** (108 mg, 66%, d.r. 96:4). Colorless oil. R_f(Et₂O/pentane 1:6) 0.34.

1H-NMR (400 MHz, CDCl₃): 0.72 (d, J=6.7, Me); 0.87 (d, J=7.0, Me); 1.23 (s, 'Bu); 1.93 – 2.01 (m, Me₂CH); 2.27 (dd, J=16.9, 4.5, 1 H, CH₂); 2.23 – 2.30 (m, 1 H, CH₂); 2.54 (dd, J=16.9, 10.5, 1 H, CH₂); 2.51 – 2.57 (m, 1 H, CH₂); 3.95 – 4.02 (m, CH₂CH); 5.06 – 5.13 (m, CH₂=); 5.45 (d, J=2.8, CHN); 5.76 – 5.86 (m, CH=); 7.25 – 7.51 (m, 8 arom. H); 7.44 – 7.49 (m, 2 arom. H).

13C-NMR (100 MHz, CDCl₃): 16.18, 22.00, 28.03 (Me); 30.63 (CH); 36.18, 36.90 (CH₂); 39.64, 64.61 (CH); 80.55, 89.52 (C); 118.09 (CH₂); 126.14, 126.33, 128.22, 128.73, 128.89, 129.25, 135.04 (CH); 139.03, 142.97, 152.97, 171.15, 175.04 (C). MALDI-MS: 500.2 (36.73), 444.2 (100.00). HR-MALDI-MS: 500.2447 ([C₂₉H₃₅NNaO₅]⁺; calc. 500.2407).

(R)-3-((R)-2-{[(Benzyloxy)carbonyl]methyl]-3-[(tert-butoxy)carbonyl]-1-oxopropyl)-4-(1-methylethyl)-5,5-diphenyloxazolidin-2-one (6). Transformation of **3** (3.2 g, 7.32 mmol) according to GP 3, followed by FC (Et₂O/pentane 1:4) afforded, beside the recovery of unreacted starting material (0.8 g, 25%), compound **6** (2.43 g, 57%). Colorless oil. $R_{\rm f}$ (Et₂O/pentane 1:4) 0.24. $[a]_{\rm L}^{\rm rt}$ = +124.5 (c = 0.80, CHCl₃). IR (CHCl₃): 3015w, 2974w, 2933w, 1780s, 1728s, 1703m, 1451w, 1364m, 1318w, 1330m, 1159s, 1051w. ¹H-NMR (400 MHz, CDCl₃): 0.75 (d, J = 6.8, Me); 0.86 (d, J = 70, Me); 1.30 (s, Bu); 1.91 – 1.99 (m, Me₂CH); 2.23 (dd, J = 16.5, 6.5, 1 H, CH₂); 2.37 (dd, J = 16.5, 7.8, 1 H, CH₂); 2.62 (dd, J = 16.5, 6.6, 1 H, CH₂); 2.90 (dd, J = 16.5, 7.3, 1 H, CH₂); 4.32 – 4.39 (m, CH₂CH); 5.11 (d, J = 7.9, PhCH₂); 5.38 (d, J = 2.9, CHN); 7.24 – 7.48 (m, 15 arom. H). ¹³C-NMR (100 MHz,

CDCl₃): 16.04, 21.65, 27.92 (Me); 30.11 (CH); 35.65, 36.05 (CH₂); 64.82 (CH); 66.60 (CH₂); 80.72, 89.42 (C); 125.68, 125.80, 125.96, 127.91, 128.23, 128.25, 128.34, 128.54, 128.57, 128.85 (CH); 135.72, 138.29, 142.30, 152.59, 169.83, 170.86, 173.71 (C). MALDI-MS: 608.2 (44.44), 552.2 (100.00). Anal. calc. for $C_{35}H_{39}NO_7$ (585.70): C 71.78, H 6.71, N 2.39; found: C 71.95, H 6.62, N 2.44.

 $(R) - 3 - ((R) - 2 - \{[(Benzyloxy) carbonyl] methyl] - 4 - [(tert-butoxy) carbonyl] - 1 - oxobutyl) - 4 - (1 - methylethyl) - 5,5 - diphenyloxazolidin - 2 - one (7). Transformation of$ **4**(4.3 g, 9.52 mmol) according to <math>GP 3, followed by FC (Et₂O/pentane 1 : 4), afforded **7** (3.14 g, 55%). Colorless oil. $R_{\rm f}$ (Et₂O/pentane 1 : 4) 0.33. $[\alpha]_{\rm L}^{\rm ph} = + 112.8$ (c = 0.68, CHCl₃). IR (CHCl₃): 2930w, 1781s, 1727s, 1598m, 1451w, 1354m, 1313w, 1251w, 1239m, 1158s, 1149w. $^{\rm 1}$ H-NMR (400 MHz, CDCl₃): 0.74 (d, J = 6.7, Me); 0.81 (d, J = 7.0, Me); 1.31 (s, Bu); 1.41 - 1.68 (m, 2 CH₂); 1.87 - 1.95 (m, Me₂CH); 2.43 (dd, J = 16.9, 4.4, 1 H, CH₂); 2.88 (dd, J = 16.9, 10.3, 1 H, CH₂); 4.14 - 4.21 (m, CH₂CH); 5.03 (d, J = 10.3, PhCH₂); 5.63 (d, J = 3.0, CHN); 7.15 - 7.38 (m, 11 arom. H); 7.43 - 7.44 (m, 2 arom. H). 13 C-NMR (100 MHz, CDCl₃): 16.02, 21.52 (Me); 27.03 (CH₂); 28.03 (Me); 30.02 (CH); 31.87, 36.37 (CH₂); 37.92, 65.29 (CH); 66.49 (CH₂); 80.20, 89.24 (C); 125.44, 125.78, 127.95, 128.18, 128.20, 128.43, 128.52, 128.70, 128.92 (CH); 135.78, 137.94, 142.46, 152.75, 171.38, 171.54, 174.71 (C). HR-MALDI-MS: 622.2775 ([C₃₆H₄₁NNaO₇]+; calc. 622.2781).

(R)-3-{(R)-}-((tert-Butoxy) carbonyl]methyl]-4-[4-(1-methylethyl)-2-oxo-5,5-diphenyloxazolidin-3-yl]-4-oxo-butanoic acid (8). Transformation of 6 (2.4 g, 4.1 mmol) according to GP 4, followed by FC (Et₂O/pentane/AcOH 10:20:0.1), afforded 8 (1.67 g, 82%). White foam. $R_{\rm f}$ (Et₂O/pentane/AcOH 10:20:0.1) 0.31. [α] $_{\rm f}^{\rm LL}$ = +148.0 (c = 1.00, CHCl₃). IR (CHCl₃): 2980w, 1780s, 1725s, 1601w, 1494w, 1450w, 1368m, 13196m, 1176m, 1153m, 1052w, 990w, 942w, 844w, 638w. ¹H-NMR (400 MHz, CDCl₃): 0.75 (d, J = 6.7, Me); 0.86 (d, J = 7.0, Me); 1.30 (s, 'Bu); 1.91 – 1.99 (m, Me₂CH); 2.24 (dd, J = 16.4, 6.5, 1 H, CH₂); 2.36 (dd, J = 16.7, 6.6, 1 H, CH₂); 2.86 (dd, J = 16.7, 7.0, 1 H, CH₂); 4.25 – 4.32 (m, CH₂CH); 5.38 (d, J = 3.0, CHN); 7.24 – 7.46 (m, 10 arom. H). ¹³C-NMR (100 MHz, CDCl₃): 16.05, 21.63, 27.91 (Me); 30.08 (CH); 36.00 (CH₂); 36.55, 64.81 (CH); 80.88, 89.51 (C); 125.82, 125.99, 127.91, 128.34, 128.57, 128.85 (CH); 138.24, 142.25, 152.65, 170.07, 173.72, 176.64 (C). MALDI-MS: 518.2 (68.93), 462.1 (100.00). Anal. calc. for C_{28} H₃₃NO $_{7}$ (495.57): C 67.86, H 6.71, N 2.83; found: C 67.74, H 6.54, N 2.76.

(R)-5-[(tert-Butoxy)carbonyl]-3-[(R)-4-(1-methylethyl)-2-oxo-5,5-diphenyloxazolidine-3-carbonyl]pentanoic Acid (9). Transformation of **7** (2.67 g, 4.45 mmol) according to GP 4, followed by FC (Et₂O/pentane/AcOH 10:20:0.1), afforded **9** (2.02 g, 89%). White foam. $R_{\rm f}$ (Et₂O/pentane/AcOH 10:20:0.1) 0.22. [a]_b^{1.5} = +121.5 (c = 0.95, CHCl₃). IR (CHCl₃): 3015w, 2974m, 2933w, 1779s, 1718s, 1451w, 1390w, 1364m, 1318w, 1256w, 1236w, 1174m, 1149w. ¹H-NMR (400 MHz, CD₃OD): 0.78 (d, J = 6.7, Me); 0.92 (d, J = 7.0, Me); 1.39 (s, Bu); 1.39 – 1.46 (m, CH₂); 1.60 – 1.65 (m, CH₂); 2.01 – 2.08 (m, Me₂CH); 2.45 (dd, J = 17.0, 4.4, 1 H, CH₂); 2.78 (dd, J = 17.0, 10.6, 1 H, CH₂); 4.14 – 4.21 (m, CH₂CH); 5.52 (d, J = 2.7, CHN); 7.25 – 7.31 (m, 2 arom. H); 7.34 – 7.39 (m, 4 arom. H); 7.48 – 7.52 (m, 2 arom. H); 7.57 – 7.60 (m, 2 arom. H). ¹³C-NMR (100 MHz, CDCl₃): 16.34, 21.93 (Me); 28.30 (CH₂); 28.36 (Me); 31.46 (CH); 32.98, 37.13 (CH₂); 39.29, 66.74 (CH); 81.64, 90.57 (C); 126.48, 126.82, 129.08, 129.80, 130.05 (CH); 139.54, 144.44, 154.39, 173.48, 174.99, 176.44 (C). HR-MALDI-MS: 532.2306 ([C₂₉H₃₅NNaO₇]⁺; calc. 532.2312).

tert-*Butyl* (\$)-3-([[(Benzyloxy)carbonyl]amino]methyl)-4-[(R)-4-(1-methylethyl)-2-oxo-5,5-diphenyloxa-zolidin-3-yl]-4-oxobutanoate (**10**). Transformation of **8** (2.0 g, 4.0 mmol) according to *GP* 5, followed by FC (Et₂O/pentane 1:3 \rightarrow 1:2), afforded **10** (1.84 g, 76%). White solid. M.p. 132 – 134°. $R_{\rm f}$ (Et₂O/pentane 1:2) 0.28. [a] $_{\rm b}^{\rm i-}$ = +123.0 (c = 1.00, CHCl₃). IR (CHCl₃): 3438w, 3008w, 2966w, 2992w, 1777s, 1722s, 1516m, 1450m, 1394w, 1368m, 1248m, 1154m, 1052w, 990w, 845w. $^{\rm i}$ H-NMR (400 MHz, CDCl₃): 0.69 (d, J = 6.7, Me); 0.84 (d, J = 7.0, Me); 1.26 (s, Bu); 1.91 – 1.99 (m, Me₂CH); 2.30 (dd, J = 16.8, 4.6, 1 H, CH₂); 2.65 (dd, J = 16.8, 9.9, 1 H, CH₂); 3.44 – 4.60 (m, CH₂); 4.09 – 4.13 (m, CH₂CH); 5.07 (s, PhCH₂); 5.23 (br. s, NH); 5.40 (d, J = 3.0, CHN); 7.23 – 7.46 (m, 15 arom. H). $^{\rm i3}$ C-NMR (100 MHz, CDCl₃): 16.01, 21.79, 27.90 (Me); 30.01 (CH); 34.81 (CH₂); 40.07 (CH); 42.80 (CH₂); 64.87 (CH); 66.81 (CH₂); 80.69, 89.67 (C); 125.79, 125.82, 127.93, 128.09, 128.16, 128.36, 128.48, 128.54, 128.55 (CH); 136.42, 138.25, 142.08, 153.18, 156.26, 170.13, 173.36 (C). MALDI-MS: 639.2 (3.07), 623.3 (20.11), 567.2 (100.00), 286.1 (44.30). Anal. calc. for C₃₅H₄₀N₂O₇ (600.70): C 69.98, H 6.71, N 4.66; found: C 69.94, H 6.80, N 4.56.

tert-Butyl (S)-4-([[(Benzyloxy)carbonyl]amino]methyl)-5-[(R)-4-(1-methylethyl)-2-oxo-5,5-diphenyloxazolidin-3-yl)-5-oxopentanoate (11). Transformation of 9 (5.94 g, 11.6 mmol) according to GP 5, followed by FC (Et₂O/pentane 1:3) \rightarrow 1:2), afforded 11 (5.4 g, 75%). Colorless oil. $R_{\rm f}$ (Et₂O/pentane 1:2) 0.32. [a] $_{\rm L}^{\rm pl}$ = +75.8 (c = 0.85, CHCl₃). IR (CHCl₃): 3015m, 2974w, 2933w, 1780s, 1718s, 1513m, 1451m, 1364m, 1313m, 1231s, 1176s, 1154s, 1050w, 990w. $^{\rm th}$ -NMR (400 MHz, (D₆)DMSO): 0.61 (d, J = 6.7, Me); 0.85 (d, J = 7.0, Me); 1.32 (s, 'Bu); 1.37 – 1.48 (m, 4 H, 2 CH₂); 2.00 – 2.08 (m, Me₂CH); 3.22 – 3.29 (m, CH₂N); 3.78 – 3.82 (m, CH₂CH); 4.93 – 5.02 (m, PhCH₂); 5.58 (d, J = 2.4, CHN); 7.24 – 7.41 (m, 12 H, 11 arom. H, NH); 7.57 (d, J = 7.2, 2 arom. H), 7.64 (d,

 $J = 7.4, 2 \text{ arom. H}). \ ^{13}\text{C-NMR} \ (100 \text{ MHz}, (D_6)\text{DMSO}): 15.27, 20.90 \ (\text{Me}); 24.48 \ (\text{CH}_2); 27.56 \ (\text{Me}); 29.21 \ (\text{CH}); 31.01 \ (\text{CH}_2); 41.62 \ (\text{CH}); 41.99 \ (\text{CH}_2); 64.80 \ (\text{CH}); 65.11 \ (\text{CH}_2); 79.39, 88.65 \ (\text{C}); 124.74, 125.16, 127.54, 127.61, 127.71, 128.17, 128.26, 128.36, 128.63 \ (\text{CH}); 136.99, 137.80, 142.86, 152.35, 156.01, 170.87, 173.14 \ (\text{C}). \ HR-MALDI-MS: 637.2884 \ ([C_{36}H_{42}N_2NaO_7]^+; calc. 637.2890). \ Anal. calc. for $C_{36}H_{42}N_2O_7$ \ (614.73): C 70.34, H 6.89, N 4.56; found: C 70.30, H 6.75, N 4.62.$

(S)-2-([[(Benzyloxy)carbonyl]amino]methyl)-3-[(tert-butoxy)carbonyl]propanoic Acid (Z- β^2 hAsp-(O'Bu)-OH; **12**). Transformation of **10** (3.0 g, 5.0 mmol) according to GP 6, followed by FC (Et₂O/pentane/AcOH 20:10:0.1), afforded **12** (1.5 g, 89%). Colorless oil. R_f (Et₂O/pentane/AcOH 20:10:0.1) 0.40. [α]_D^{L.} = +35.4 (c = 0.85, CHCl₃). IR (CHCl₃): 3436m, 3036m, 2974m, 1718s, 1513s, 1451m, 1364m, 1148s. ¹H-NMR (400 MHz, CD₃OD): 1.43 (s, ¹Bu); 2.43 (dd, J = 16.6, 5.2, 1 H, CH₂); 2.55 (dd, J = 16.6, 8.8, 1 H, CH₂); 2.90 – 2.96 (m, CH₂CH); 3.28 – 3.38 (m, 1 H, CH₂N); 3.43 (dd, J = 13.9, 6.3, 1 H, CH₂N); 5.07 (s, PhCH₂); 7.27 – 7.34 (m, 5 arom. H). ¹³C-NMR (100 MHz, CD₃OD): 28.32 (Me); 36.04, 43.00 (CH₂); 43.41 (CH); 67.55 (CH₂); 82.07 (C), 128.82, 129.00, 129.49 (CH); 138.38, 158.93, 172.75, 176.56 (C). HR-MALDI-MS: 360.1418 ([C₁₇H₂₃NNaO₆]⁺; calc. 360.1423).

(S)-2-([[(Benzyloxy)carbonyl]amino]methyl)-4-[(tert-butoxy)carbonyl]butanoic Acid (Z- β^2 hGlu(O'Bu)-OH; **13**). Transformation of **11** (2.7 g, 4.39 mmol) according to GP 6, followed by FC (Et₂O/pentane/AcOH 20:10:0.1), afforded acid **13** (1.5 g, 93%). Colorless oil. R_f (Et₂O/pentane/AcOH 20:10:0.1) 0.35. [α]_L^{T-1} = +2.3 (c = 1.03, CHCl₃). IR (CHCl₃): 3446w, 2974w, 1718s, 1513m, 1451w, 1369m, 1149m. ¹H-NMR (400 MHz, (D₆)DMSO): 1.31 (s, 'Bu); 1.52 – 1.66 (m, CH₂); 2.04 – 2.21 (m, CH₂CO); 2.36 – 2.40 (m, CH₂CH); 2.96 – 3.03 (m, 1 H, CH₂N); 3.11 – 3.18 (m, 1 H, CH₂N); 4.94 (s, PhCH₂); 7.21 – 7.31 (m, 5 arom. H); 12.25 (br. s, CO₂H). ¹³C-NMR (100 MHz, (D₆)DMSO): 24.26 (CH₂); 27.63 (Me); 32.28, 41.82 (CH₂); 44.26 (CH); 65.12 (CH₂); 79.53 (C); 127.54, 127.63, 128.23 (CH); 137.06, 156.07, 171.54, 174.77 (C). HR-MALDI-MS: 374.1574 ([C₁₈H₂₅NNaO₆]⁺; calc. 374.1580). Anal. calc. for C₁₈H₂₅NO₆ (351.39): C 61.53, H 7.17, N 3.99; found: C 61.32, H 7.13, N 4.00.

(S)-3-[(tert-Butoxy)carbonyl]-2-([[(9H-fluoren-9-yl)carbonyl]amino]methyl)propanoic Acid (Fmocβ²hAsp(OʻBu)-OH; **14**). The Z-protected acid **12** (1 g, 2.96 mmol) in MeOH (0.02м) was hydrogenated according to GP 7. The resulting product was Fmoc-protected according to GP 8. FC (Et₂O/pentane/AcOH 20:10:0.1) of the crude product afforded **14** (1.18 g, 94%). White solid. M.p. 109–113°. $R_{\rm f}$ (Et₂O/pentane/AcOH 20:10:0.1) 0.19. [α] $_{\rm i}^{\rm id}$ = +1.4 (c = 1.20, CHCl₃). IR (CHCl₃): 3450w, 3015m, 2981m, 1723s, 1515s, 1450m, 1369m, 1236s, 1153s. $^{\rm ih}$ -NMR (400 MHz, (D₆)DMSO): 1.38 (s, $^{\rm ih}$ Bu); 2.30–2.45 (m, COCH₂); 2.73–2.80 (m, CHCl₂N); 3.08–3.15 (m, 1 H, CH₂N); 3.25–3.30 (m, 1 H, CH₂N); 4.21 (t, J = 70, CHCH₂O); 4.30 (d, J = 73, CHCH₂O); 7.31–7.44 (m, 4 arom. H, NH); 7.68 (d, J = 7.4, 2 arom. H), 7.88 (d, J = 7.5, 2 arom. H); 12.38 (br. s, CO₂H). $^{\rm is}$ C-NMR (100 MHz, (D₆)DMSO): 27.60 (Me); 34.40, 41.41 (CH₂); 41.62, 46.64 (CH); 65.28 (CH₂); 79.76 (C); 120.01, 125.08, 126.94, 127.50 (CH); 140.63, 143.76, 156.09, 170.53, 173.97 (C). HR-MALDI-MS: 448.1731 ([C₂₄H₂₇NNaO₆] $^{+}$; calc. 448.1736). Anal. calc. for C₂₄H₂₇NO₆ (425.47): C 67.75, H 6.40, N 3.29; found: C 67.84, H 6.39, N 3.34.

(S)-4-[(tert-Butoxy)carbonyl]-2-([[(9H-fluoren-9-yl)carbonyl]amino]methyl)butanoic Acid (Fmocβ²hGlu(OʻBu)-OH; **15**). The Z-protected acid **13** (0.2 g, 0.57 mmol) in MeOH (0.02M) was hydrogenated according to GP 7. The resulting product was Fmoc-protected according to GP 8. FC (Et₂O/pentane/AcOH 20:10:0.1) of the crude product afforded **15** (225 mg, 90%). Colorless oil. $R_{\rm f}$ (Et₂O/pentane/AcOH 20:10:0.1) 0.18. [α] $_{\rm B}^{\rm id}$ = +1.8 (c = 0.85, CHCl₃). IR (CHCl₃): 3445w, 3010m, 2977w, 1720s, 1514m, 1448m, 1369m, 1225m, 1150s. $^{\rm i}$ H-NMR (400 MHz, (D₆)DMSO): 1.38 (s, 'Bu); 1.63 – 1.73 (m, COCH₂CH₂); 2.13 – 2.27 (m, COCH₂); 2.44 – 2.49 (m, CH₂CH); 3.06-3.10 (m, 1 H, CH₂N); 3.20 – 3.25 (m, 1 H, CH₂N); 4.20 (t, J = 5.5, CHCH₂O); 4.27 (d, J = 4.7, CHCH₂O); 7.31 – 7.45 (m, 4 arom. H, NH); 7.69 (d, J = 6.0, 2 arom. H), 7.88 (d, J = 6.0, 2 arom. H); 12.32 (br. s, CO₂H). 13 C-NMR (100 MHz, (D₆)DMSO): 24.25 (CH₂); 27.64 (Me); 32.29, 41.78 (CH₂); 44.24, 46.62 (CH); 65.33 (CH₂); 79.54 (C); 119.99, 125.10, 126.95, 127.50 (CH); 140.62, 143.78, 156.05, 171.55, 174.79 (C). HR-MALDI-MS: 462.1887 ([C₂₅H₂₉NNaO₆]+; calc. 462.1893). Anal. calc. for C₂₅H₂₉NO₆ (439.50): C 68.32, H 6.65, N 3.19; found: C 68.33, H 6.81, N 3.19.

1-Benzyl 4-(tert-Butyl) (S)-2-([(Benzyloxy)carbonyl]amino]methyl)butanedioate (Z-β²hAsp(OʻBu)-OBn; **16**). Transformation of **12** (2.7 g, 8.0 mmol) according to GP 9, followed by FC (Et₂O/pentane 1:2), afforded **16** (3.11 g, 91%). Colorless oil. R_t (Et₂O/pentane 2:1) 0.34. [a] $_D^{\text{LL}} = +1.6$ (c = 1.03, CHCl₃). IR (CHCl₃): 3446w, 3015m, 2974w, 1723s, 1513m, 1456w, 1364m, 1251m, 1149s. ¹H-NMR (400 MHz, CD₃OD): 1.37 (s, 'Bu); 2.45 (dd, J = 16.8, 4.8, 1 H, COCH₂); 2.60 (dd, J = 16.8, 9.4, 1 H, COCH₂); 2.97 –3.03 (m, CH₂CH); 3.33 –3.43 (m, CH₂N); 5.04 –5.13 (m, 2 PhCH₂); 7.27 –7.36 (m, 10 arom. H). ¹³C-NMR (100 MHz, CD₃OD): 28.25 (Me); 35.85, 43.03 (CH₂); 45.58 (CH); 67.53, 67.75 (CH₂); 82.13 (C); 128.80, 128.97, 129.17, 129.24, 129.45,

129.51 (CH); 138.28, 137.39, 158.83, 172.44, 174.56 (C). HR-MALDI-MS: 450.1887 ([C₂₄H₂₉NNaO₆]⁺; calc. 450.1893). Anal. calc. for C₂₄H₂₉NO₆ (427.49): C 67.43, H 6.84, N 3.28; found: C 67.18, H 6.84, N 3.53.

1-Benzyl 5-(tert-Butyl) (S)-2-([[(Benzyloxy)carbonyl]amino]methyl)pentanedioate (Z-β²hGlu(OʻBu)-OBn; 17). Transformation of 13 (3.0 g, 8.5 mmol) according to GP 9, followed by FC (Et₂O/pentane 1:2), afforded 17 (3.5 g, 93%). Colorless oil. $R_{\rm f}$ (Et₂O/pentane 2:1) 0.26. [α]_D·· = +5.7 (c = 0.58, CHCl₃). IR (CHCl₃): 3446w, 3026w, 2995w, 1718s, 1513m, 1456w, 1364w, 1236m, 1149m. ¹H-NMR (400 MHz, (D₆)DMSO): 1.37 (s, Bu); 1.67 – 1.73 (m, COCH₂CH₂); 2.10 – 2.25 (m, COCH₂); 2.57 – 2.65 (m, CH₂CH); 3.11 – 3.18 (m, 1 H, CH₂N); 3.22 – 3.29 (m, 1 H, CH₂N); 5.00 – 5.10 (m, 2 PhCH₂); 7.28 – 7.36 (m, 10 arom. H); 7.44 (t, J = 5.9, NH). ¹³C-NMR (100 MHz, (D₆)DMSO): 24.24 (CH₂); 27.61 (Me); 32.04, 41.90 (CH₂); 44.33 (CH); 65.20, 65.64 (CH₂); 79.53 (C); 127.56, 127.65, 127.75, 127.87, 128.21, 128.27 (CH); 136.01, 137.00, 156.08, 171.42, 173.07 (C). HR-MALDI-MS: 464.2044 ([C₂₅H₃₁NNaO₆]⁺; calc. 464.2049). Anal. calc. for C₂₅H₃₁NO₆ (441.52): C 68.01, H 7.08, N 3.17; found: C 61.12, H 6.97, N 3.29.

Benzyl (S)-3-(Aminocarbonyl)-2-([[(benzyloxy)carbonyl]amino]methyl)propanoate (Z-β²hAsn-OBn; **18**). The diester **16** (0.55 g, 1.29 mmol) was 'Bu deprotected according to *GP 10*. The acid obtained was then converted to **18** according to *GP 11*. Recrystallization of the crude product yielded **18** (345 mg, 72%). White solid. M.p. 130 – 131°. R_t (AcOEt/Et₂O 2:1) 0.2. [α]₁₅·l = -18.7 (c = 0.92, CHCl₃). IR (CHCl₃): 3518w, 3426w, 3405w, 3015w, 2954w, 1723s, 1682s, 1595w, 1513m, 1451w, 1226s, 1174w. ¹H-NMR (400 MHz, (D₆)DMSO): 2.28 (dd, J = 15.8, 5.1, 1 H, COCH₂); 2.44 (dd, J = 15.8, 9.0, 1 H, COCH₂); 2.90 – 2.97 (m, CH₂CH); 3.16 – 3.29 (m, CH₂N); 4.97 – 5.07 (m, 2 PhCH₂); 6.82 (br. s, 1 H, NH₂); 7.28 – 7.37 (m, 11 H, 10 arom. H, NH₂); 7.42 (t, J = 6.0, NH). ¹³C-NMR (100 MHz, (D₆)DMSO): 34.17, 41.64 (CH₂); 41.83 (CH); 65.23, 65.50 (CH₂); 127.51, 127.59, 127.67, 127.70, 128.21, 128.23 (CH); 136.11, 136.97, 156.09, 171.92, 172.90 (C). HR-MALDI-MS: 393.1421 ([C₂₀H₂₂N₂NaO₅]⁺; calc. 393.1427). Anal. calc. for C₂₀H₂₂N₂O₅ (370.40): C 64.85, H 5.99, N 7.56; found: C 64.83, H 5.95, N 7.43.

Benzyl (S)-4-(Aminocarbonyl)-2-([[(benzyloxy)carbonyl]amino]methyl)butanoate (Z-β²hGln-OBn; 19). The diester 17 (3.5 g, 7.93 mmol) was 'Bu deprotected according to GP 10. The acid obtained was then converted to 19 according to GP 11. Recrystallization of the crude product yielded 19 (2.63 g, 83%). White solid. M.p. $112-113^\circ$. R_1 (EtOAc/Et₂O 2:1) 0.28. [α]_D·= +2.5 (c=0.95, CHCl₃). IR (CHCl₃): 3528w, 3446w, 3405w, 3036w, 3005w, 2964w, 1718s, 1682s, 1590w, 1513m, 1451w, 1226m, 1164w. ¹H-NMR (400 MHz, (D₆)DMSO): 1.66 – 1.73 (m, COCH₂CH₂); 1.99 – 2.12 (m, COCH₂); 2.56 – 2.63 (m, CH₂CH); 3.13 – 3.29 (m, CH₂N); 5.00 (s, PhCH₂); 5.06 (s, PhCH₂); 6.73 (s, 1 H, NH₂); 7.27 – 7.37 (m, 10 arom. H); 7.42 (t, t = 5.9, NH). ¹³C-NMR (100 MHz, (D₆)DMSO): 24.75, 32.20, 42.01 (CH₂); 44.79 (CH); 65.18, 65.58 (CH₂); 127.56, 127.65, 127.71, 127.83, 128.23, 128.28 (CH); 136.02, 137.00, 156.06, 173.27 (C). HR-MALDI-MS: 407.1577 ([C₂₁H₂₄N₂NaO₅]⁺; calc. 407.1583). Anal. calc. for C₃₁H₂₄N₃N₅(384.43): C 65.61, H 6.29, N 7.29; found: C 65.67, H 6.50, N 7.37.

Benzyl (S)-2-([[(Benzyloxy)carbonyl]amino]methyl)-3-[[triphenylmethyl)amino]carbonyl]propanoate (Z-β²hAsn(Trt)-OBn; **20**). Treatment of **18** (0.3 g, 0.81 mmol) according to GP 12, followed by FC (Et₂O/pentane 3:1), afforded **20** (421 mg, 85%). White solid. M.p. $104-105^\circ$. R_f (Et₂O/pentane 3:1) 0.29. [α] $_D^{\text{Li}}=-20.8$ (c=1.15, CHCl₃). IR (CHCl₃): 3434w, 3028w, 3020w, 1721s, 1513s, 1492s, 1446w, 1236m, 1195w, 1169w. $^{\text{Li}}$ H-NMR (400 MHz, (D₆)DMSO): 2.45 (dd, J=15.6, 5.2, 1 H, COCH₂); 2.75 (dd, J=15.6, 8.9, 1 H, COCH₂); 2.83 – 2.90 (m, CH₂CH); 3.12 – 3.26 (m, CH₂N); 4.96 – 5.05 (m, 2 PhCH₂); 7.14 – 7.34 (m, 25 arom. H); 7.38 (t, J=6.1, NH); 8.69 (s, NHTrt). 13 C-NMR (100 MHz, (D₆)DMSO): 35.20, 41.71 (CH₂); 41.88 (CH); 65.25, 65.60 (CH₂); 69.24 (C); 126.21, 127.30, 127.56, 127.62, 127.68, 127.72, 128.03, 128.22, 128.42 (CH); 136.01, 136.95, 144.69, 156.08, 169.75, 172.79 (C). HR-MALDI-MS: 635.2516 ([C₃₀H₃₆N₂NaO₅]⁺; calc. 635.2522). Anal. calc. for C₃₀H₃₆N₂O₅ (612.71): C 76.45, H 5.92, N 4.57; found: C 76.27, H 6.02, N 4.57.

Benzyl (S)-2-([[(Benzyloxy)carbonyl]amino]methyl)-4-[[(triphenylmethyl)amino]carbonyl]butanoate (Z-β²hGln(Trt)-OBn; **21**). Treatment of **19** (2.66 g, 6.9 mmol) according to GP 12, followed by FC (Et₂O/pentane 3:1), afforded **21** (3.81 g, 88%). White solid. M.p. 91 – 92°. $R_{\rm f}$ (Et₂O/pentane 3:1) 0.3. [α] $_{\rm i}^{\rm pt}$ = −4.6 (c = 0.90, CHCl₃). IR (CHCl₃): 3536w, 3067w, 3035w, 3005w, 1718s, 1682m, 1512s, 1492s, 1446w, 1235m, 1164w. $^{\rm t}$ H-NMR (400 MHz, (D₆)DMSO): 1.63 – 1.68 (m, COCH₂CH₂); 2.27 – 2.30 (m, COCH₂); 2.52 – 2.59 (m, CH₂CH); 3.12 – 3.25 (m, CH₂N); 4.99 (m, PhCH₂); 5.07 (m, PhCH₂); 7.14-7.32 (m, 25 arom. H); 7.33 (m, J = 2.4, NH) 7.23 (m, NHTrt). $^{\rm t}$ 3-C-NMR (100 MHz, (D₆)DMSO): 24.89, 33.09, 41.91 (CH₂); 44.73 (CH); 65.18, 65.56 (CH₂); 69.10 (C); 126.19, 127.32, 127.56, 127.64, 127.67, 127.81, 128.22, 128.27 (CH); 136.04, 136.98, 144.79, 156.07, 171.09, 173.29 (C). HR-MALDI-MS: 649.2673 ([C₄₀H₃₈N₂NaO₅] $^+$; calc. 649.2679). Anal. calc. for C₄₀H₃₈N₂O₅ (626.74): C 76.66, H 6.11, N 4.47; found: C 76.65, H 6.16, N 4.41.

(S)-2-([(9H-Fluoren-9-yl)carbonyl]amino]methyl)-3-(<math>[(triphenylmethyl)amino]carbonyl]propanoic Acid (Fmoc- β^2 hAsn(Trt)-OH; **22**). Hydrogenation of **20** (0.39 g, 0.64 mmol) in MeOH (0.02M) according to GP 7 afforded the crude H- β^2 hAsn(Trt)-OH, which was Fmoc-protected according to GP 8 to give, after FC (AcOEt/

pentane/AcOH 20:10:0.1), the acid **22** (0.36 g, 93%). White foam. $R_{\rm f}$ (AcOEt/pentane/AcOH 20:10:0.1) 0.4. [α]_D: $^{\rm L}$ = -29.6 (c = 1.35, CHCl₃). IR (CHCl₃): 3436w, 3056w, 3005w, 2995w, 1718s, 1672w, 1625w, 1513s, 1492s, 1446w, 1231w. $^{\rm L}$ H-NMR (400 MHz, (D₆)DMSO): 2.36 (dd, J = 15.2, 4.8, 1 H, COCH₂); 2.67 (dd, J = 15.2, 9.1, 1 H, COCH₂); 2.72 - 2.75 (m, CH₂CH); 3.06 - 3.13 (m, 1 H, CH₂N); 3.18 - 3.26 (m, 1 H, CH₂N); 4.19 - 4.22 (m, CHCH₂O); 4.26 - 4.28 (m, CH₂O); 5.01 (d, J = 1.8, NHCH₂); 7.15 - 7.21 (m, 9 arom. H); 7.23 - 7.42 (m, 10 arom. H); 7.68 (d, J = 7.4, 2 arom. H); 7.88 (d, J = 7.5, 2 arom. H); 8.64 (s, NHTrt); 12.27 (br. s, CO₂H). 13 C-NMR (100 MHz, (D₆)DMSO): 34.43, 41.66 (CH₂); 41.80, 46.60 (CH); 65.36 (CH₂); 69.20 (C); 120.00, 125.10, 126.20, 126.97, 127.30, 127.51, 127.60, 127.65, 128.23, 128.46 (CH); 137.03, 140.60, 143.74, 143.79, 144.79, 156.05, 170.07, 174.53, 174.56 (C). HR-MALDI-MS: 633.2366 ([C₃₀H₃₄N₂NaO₅]⁺; calc. 633.2360).

(S)-2-([[(9H-Fluoren-9-yl)carbonyl]amino]methyl)-4-[[(triphenylmethyl)amino]carbonyl]butanoic Acid (Fmoc-β²hGln(Trt)-OH; **23**). Hydrogenation of **21** (0.25 g, 0.4 mmol) in MeOH (0.02M) according to *GP 7* afforded the crude H-β²hGln(Trt)-OH, which was Fmoc-protected according to *GP 8* to give, after FC (AcOEt/pentane/AcOH 20:10:0.1), the acid **23** (0.23 g, 92%). White foam. $R_{\rm f}$ (AcOEt/pentane/AcOH 20:10:0.1) 0.22. [α] $_{\rm b}$ ¹ = $_{\rm f}$ = $_{\rm f}$ = 0.95, CHCl $_{\rm f}$). IR (CHCl $_{\rm f}$): 3436w, 3024w, 2974w, 1713s, 1513s, 1492s, 1446m, 1236m, 1149w, 1103w. $_{\rm f}$ H-NMR (400 MHz, (D $_{\rm f}$)DMSO): 1.59 – 1.66 (m, COCH $_{\rm f}$ CH $_{\rm f}$); 2.27 – 2.32 (m, COCH $_{\rm f}$); 2.41 – 2.45 (m, CH $_{\rm f}$ CH); 3.04 – 3.11 (m, 1 H, CH $_{\rm f}$ N); 3.16 – 3.23 (m, 1 H, CH $_{\rm f}$ N); 4.18 – 4.21 (m, CHCH $_{\rm f}$ O); 4.24 – 4.26 (m, CH $_{\rm f}$ O); 7.15 – 7.34 (m, 17 arom. H, NH); 7.40 (t, J = 7.4, 3 arom. H); 7.67 (d, J = 7.4, 2 arom. H); 7.88 (d, J = 7.5, 2 arom. H); 8.57 (s, NHTrt); 12.26 (br. s, CO $_{\rm f}$ H). $_{\rm f}$ C-NMR (100 MHz, (D $_{\rm f}$ DMSO): 25.00, 33.36, 41.81 (CH $_{\rm f}$); 44.67, 46.59 (CH); 65.35 (CH $_{\rm f}$); 69.08 (C); 119.98, 125.11, 126.17, 126.96, 127.32, 127.50, 128.23, 128.41 (CH); 140.60, 143.77, 144.83, 156.04, 171.29, 175.02 (C). HR-MALDI-MS: 647.2523 ([C $_{\rm f}$ O_{$\rm f}$ O_{$\rm f}O_{<math>\rm f}O_{<math>\rm f}O_{<math>\rm f}$ O_{$\rm f}O_{<math>\rm f}O_{<math>\rm f}O_{<math>\rm f}O_{<math>\rm f}O_{\rm f}O_{<math>\rm f}O_{<math>\rm f}O_{\rm f}O_{<math>\rm f}O_{\rm f}O_{<math>\rm f}O_{\rm f}O_{\rm f}O_{<math>\rm f}O_{\rm f}O_{<math>\rm f}O_{\rm f}O_{<math>\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{<math>\rm f}O_{\rm f}O_{\rm f}O_{<math>\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{<math>\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{<math>\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{<math>\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{<math>\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{<math>\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{\rm f}O_{\rm}}$}}}}}}}}}}}}</sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub></sub>

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